A Molecular Model for the Exobilayer Portion of the α-Subunit of the Acetylcholine Receptor with Binding Sites for Acetylcholine and Non-competitive Antagonists

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ABSTRACT A molecular model for the exobilayer portion of the α -subunit of the acetylcholine receptor is presented. Binding sites for an acetylcholine and non-competitive antagonists are indicated.

We recently presented structural assignments (1,2) for 24 transmembrane segments of the acetylcholine receptor (AChR)**, a membrane complex with two α - and one each of β -, γ - and γ -subunits.(3,4) The AChR ion channel and an acetylcholine binding site at the mouth of the ion channel are a central feature of the model (5). Our molecular model for all helices of the AChR in the bilayer accounted for 656 (28%) (6) of the 2333 amino acids in the receptor (7-12). The subunit order, α -, β -, α -, γ - and γ - around a central "pit", is favored (12), but not settled.(18)

We now present a molecular model for the exobilayer portion of the α -subunit of the AChR. Two binding sites, one for acetylcholine (ACh) and one for a non-competitive antagonist, are designated in the exobilayer region, for a total of 3 in our model.

2. THEORY AND DISCUSSION

A model for the exobilarer portion of the receptor helps us to (1) formulate labeling experiments designed to probe AChR structure (2) develop explanations for particular chemical effects (e.g., disulfide reduction) and

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^{**} AChR, acetylcholine receptor; ACh, acetylcholine; glu, glutamate; asp, aspartate; lys, lysine; DTT, dithiothreitol; SGR theory, single group rotation theory

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physiological responses (such as differences between agonists and local anesthetics) (3) open the way to theories for the operation of structurally complex receptors and ion channels (4) generalize the approach to receptor structure analysis gained from our study of the AChR bilayer helices. (1.2.5.6)

The criteria used for the design of the exobilayer model are as follows:

(1) Two binding sites. In addition to the ACh binding site near the mouth of the ion channel in the bilayer, two binding sites should appear, one for ACh near a reducible disulfide (cf.(3)), and another for noncompetitive blockers close to the \$-subunit (14). A total of three ligand sites are also implied by monoclonal antibody binding studies. (15,16) Activation of the iun channel depends upon two molecules of ACh. (17) The binding site near the disulfide may be blocked without eliminating ion channel function. (18,19)

- (2) **55A Extension from bilayer.** Electron microscopy indicates that the structural elements of the exobilayer portion of the acetylcholine receptor extend approximately 55A from the bilayer. (12)
- (3) **High antiparallel \beta-sheet content.** Raman spectroscopy indicates that antiparallel β -sheet is the most abundant form of polypeptide (34%) present in the AChR (20, 21). Most of the α -helical content of the receptor has been accounted for the bilayer α -helices.
- (4) Lysine-glutamate (aspartate) binding site The model for the bilayer binding site for ACh contained this combination of positively and negatively charged groups. The use of the (+,-) pair in the successful formulation of a model for binding and gating suggested that the same choice be made for the exobilayer sites.
- (5) One reactive thiol (SH) group per α -subunit. Dithiothreitol (DTT) could produce two reactive SH groups per disulfide reduced. However, only one is found, carefully executed affinity alkylations with bromoacetylcholine revealing one reactive thiol per α -subunit. (22) The model should thus include the possibility of forming one unreactive SH group per disulfide reduced.

We have therefore arranged strands of 17-18 amino acids (56-60A) as a folded antiparallel β -sheet (designated as strands X1-X3, X4-X6 in Fig.1) so

17	3s 174g 175	•							
172-	1	76							
171		177							
170		178							
169t		179+							
168s		180-				60 59	am	22 21 F	20rg÷
167		181		100 99-	98g	61	58am	23-	19
166-		182rg+		101a	97-	-62	57rg+	24h	18
165P		183g		102	96a	63	56	25h	17+
164rg+		184		103	95 am		55rg+	26t	16am
163-		185+		104h	94am		54	27h	15
162s	135		34h	105	93	66rg+	53am	28	14am
161-	136P		33t	106+	92	67	52t	29	13-
160P	137	188	132	107+	91	68am	51~	30-	12
159s	138-	189	131	108	90	69P	50	31	11
158	139am	190	130	109	89~	70a	49	32t	10am
157	140am	191t	129-	110	88P	71-	48am	33	9a
156	141am	1925		111=	87	72	47am	34g	8
155+	1425	1935	127	112	86	73g	46	35	7
154t	143t	194P	126s	113t	85	74g	45-	36am	6rg+
153g	144	195-	125+	114g	84-	75 ~	44-	37	5t.
152-	145+	196t	124	115+	83-	76+	43	38	4-
151	146	197P	123	116	82s	77+	42s	39am [.]	3h
150t	147g	198	122a	117	81P	78	41	40	2- 1s
149	148	199	121P	118	80	79rg+			
		200-	120P	119t					
		201							
		202							
		203							
		204h							
		205							
		206							
		207							
		208am							
		209rg+							
χ9	х8	X10	Х7	Х6	X5	Х4	Х3	X2	X1

Fig.1 Partial tertiary structure of exobilayer portion of α -subunit of AChR, the acetylcholine receptor. [+,lys; rg+,arg; -,glu or asp; 5,cys; s,ser; t,thr; h,his; g,gly; P,pro; a,ala; am,gln or asn; all hydrophobic residues [leu, ileu, val, phe, tyr, trp] unmarked (asp 30, 44, 62, 71, 83, 84, 89, 97, 99, 111, 138, 152, 166, 180, 195, 200; asn 10, 14, 16, 47, 53, 68, 94, 95, 141)

as to include half of a suitable binding site [lys and glu or asp]. (Designated as half-site A, Fig.2, upper portion) The design of the binding sites is based on single group rotation (SGR), theory. (23) The internal apposed chains (X1-X3 versus X4-X6) in the model are almost all compatible, including significant numbers of hydrophobic-hydrophobic interactions.

A lysine(-125) and an aspartate(-195) suitable for the second half-binding site (half-site B) are located near the disulfide bond. The only reasonable combination of cysteine SH groups as disulfides (proximate binding site, appropriate geometry) is 128-cys-192-cys, 142-cys-193-cys. The overall

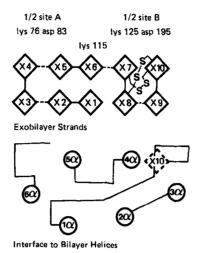


Fig.2 (a) Lower portion. The interface region between the bilayer helices and the exobilayer strands of the α -subunit. The strands are represented by parallelopipeds, the bilayer helices by circles. The strand order is shown in Fig.1. An example of a favorable interaction is that between amino acid side chains 360 val-361 ile-362 phe (4α - to 5α - connection) and 207 met-206 ile-205 phe (1α - to X18 connection). (b) Upper portion. Two half-sites are indicated for the exobilayer strands. The other half of each site would be found on the apposite exobilayer portion of the α -subunit.(shown in Scheme) The disulfide sulfur easily accessible to external reagents is that from cys-193.

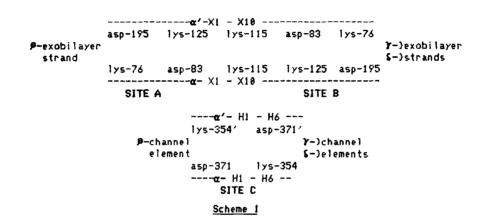
arrangement of strands X7-X10 bears a resemblance to that of the snake α-neurotoxins (24), possessing even a proline next to one disulfide, and likely to have an "open" β- pleated sheet structure. Given that the nearby binding site (lys-125, asp-195) has side chains projecting into the ion channel opening, the X7-X10 disulfide (128-cys-192-cys) would be hidden and not readily reducible, while 193-cys would carry the SH group accessible for labeling and the 142-cys SH group would be "hidden". (See Fig.2, upper portion)

The connection between the bilayer helices and the exobilayer strands is constructed so as to minimize the separation between the helices and the exobilayer polypeptide. The link is indicated schematically in the lower portion of Fig.2, with one favorable interaction among several noted in the caption.

The model for the exobilayer strands is consistent with the finding that only one disulfide per α -subunit is susceptible to reduction (25) and not inconsistent with the finding that the same subunit has seven antigenic determinants. (26) The model contains two different exobilayer binding sites. The first (A) is suitable for acetylcholine, the structure being similar to that

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of the site (C) at the ion channel mouth. (5, 6). The second (B) must be larger (because it is bounded by the Y- and S-subunits) and should be suitable for local anesthetic molecules. Affinity labeling with 5-azidotrimethoisoguin at this site links the label to the \$-subunit (14), whereas uv irradiation of AChR carrying chloropromazine leads to labeling of the α -, β - and δ subunits along with less labeling on the Y-subunit. (27) Half-sites (A and B) are indicated in the upper portion of Fig.2. The exobilayer sites (A and B) and the bilayer site (C) are shown in the Scheme below. (X = strand, H = helix)



REFERENCES

- Kosower, E.M. (1983) Biochem. Biophys. Res. Commun. 111, 1822-1826
- Kosower, E.M. (1983) FEBS Letters, 155, 245-247
 Karlin, A. (1980) in The Cell Surface and Neuronal Function, eds. Cotman, C.W., Poste, G. and Nicolson, G.L., Elsevier-North Holland, pp. 191-260 4. Lindstrom, J., Merlie, J. and Yogeeswaran, G. (1979) Biochemistry 18, 4465-4470
- 5. Kosower, E.M. (1984) (Fourth Biophysical Discussion, October, 1983) Biophys.J., In Press
- 6. Kosower, E.M., (1983) FEBS Letters, 157, 144-146
- 7. Noda, M., Takahashi, H., Tanabe, T., Toyosato, M., Furutani, Y., Hirose, T., Asai, M., Inayama, S. Miyata, T. and Numa, S. (1982) Nature 299, 793-797.
- 8. Noda, M., Takahashi, H., Tanabe, T., Toyosato, M., Kikyotani, S., Hirose, T., Asai,M., Takashima,H., Inayama,S., Miyata,T. and Numa,S. (1983) Nature 301. 251-255
- Claudio, T., Ballivet, M., Patrick, J. and Heinemann, S. (1983) Proc. Nat'1. Acad. Sci. USA 80, 1111-1115
- 18. Noda, M., Takahashi, H., Tanabe, T., Toyosato, M., Kikyotani, S., Furutani, Y., Hirose, T., Takashima, H., Inayama, S., Miyata, T. and Numa, S. (1983) Nature 302, 528-532
- 11. Devillers-Thiery,A., Giraudat,J., Bentaboulet,M. and Changeux,J-P. (1983) Proc. Nat'l. Acad. Sci. USA, 80, 2067-2071
- Stroud,R.M., Klymkowsky,M.W., LaLancette,R. Kistler,J., and Fairclough, R.H. (1982) Biophys. J. 37, 371-383.

Vol. 116, No. 1, 1983 BIOCHEMICAL AND BIOPHYSICAL RESEARCH COMMUNICATIONS

- 13. Karlin,A., Holtzman,E., Yodh,N., Lobel,P., Wall,J. and Hainfeld,J. (1983) J. Biol. Chem. 258, 6678-6681
- 14. Saitoh,T., Oswald,R. Wennogle,L.P. and Changeux,J-P. (1980) FEBS Letters 116, 30-36
- 15. Tzartos, S.J., Rand, D.E., Einarson, B.L. and Lindstrom, J.M. (1981) J. Biol. Chem. 256, 8635-8645
- 16. Watters, D. and Maelicke, A. (1983) Biochemistry 22, 1811-1819
- 17. Adams, P.R. (1981) Fed. Proc. 40, 2611-2613
- 18. Blanchard, S.G., Dunn, S.M.J. and Raftery, M.A. (1982) Biochemistry 24, 3258-3264
- Dunn, S.M.J. and Raftery, M.A. (1982) Proc. Nat'l. Acad. Sci. USA 79, 6757-6761
- 20. Chang, E.L., Yager, P., Williams, R.W. and Dalziel, A.W. (1983) Biophysical J. 41, 65a (Abstract M-AM-Pos53)
- 21. Williams, R.W. (1983) J. Mol. Biol. 166, 581-683
- 22. Wolosin, J.M., Lyddiatt, A., Dolly, J.O. and Barnard, E.A. (1980) Eur. J. Biochem. 109, 495-505
- 23. Koscwer, E.M. (1982) Abstracts, Internat'l. Symposium, Structure and Dynamics of Nucleic Acids and Proteins, San Diego, Calif. pp.52-3
- 24. Tsernoglou, D., Petsko, G.A. and Hudson, R.A. (1978) Mol. Pharm. 14, 718-716
- 25. Walker, J.W., Lukas, R.J. and McNamee, M.G. (1981) Biochemistry 20, 2191-2199
- 26. Gulick, W.J. and Lindstrom, J.M. (1983) Biochemistry 22, 3312-3320
- 27. Heidmann, T., Oswald, R.E. and Changeux, J.-P. (1983) Biochemistry 22, 3112-3127